

**Detection/characterisation of undescribed genetically  
modified bacteria by statistical analysis of high throughput  
sequencing data**

**Julie HUREL**



Supervision :

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Stéphanie BOUGEARD (statistics)  
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Viral Genetics and Biosecurity Unit (UGVB)  
ANSES Ploufragan

1. Introduction
2. Objectives
3. Data preparation
4. Calculation of the distances
5. Design of the prediction model
6. Results
7. Conclusion
8. Perspectives

# 1. Introduction

2. Objectives

3. Data préparation

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## A – Genetically Modified Organism : GMO

- ▶ Living being whose genetic material has been modified in a non-natural way
- ▶ Simplified description of the structure of a GMO



GMO

« Host » genome

Junction sequences

Insert

- ▶ Insert : often CDS(s) (coding sequence)

## A – Genetically Modified Organism : GMO

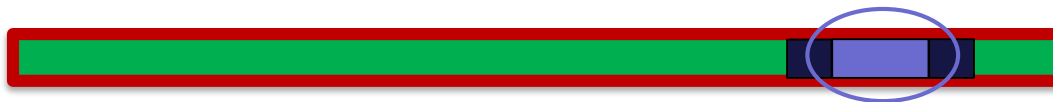
- ▶ Living being whose genetic material has been modified in a non-natural way
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## A – Genetically Modified Organism : GMO

- ▶ Living being whose genetic material has been modified in a non-natural way
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OGM

« Host » genome

Junction sequences

Insert

- ▶ Insert : often CDS(s) (coding sequence)

## B – Existing detection methods

- ▶ For known GMO
  - ▶ Methods based on protein detection or DNA detection
  - ▶ Use of qPCR : greater sensitivity and accurate quantification
  - ▶ GMOseek [Morisset et al. 2014]



*Credit: Vit Kovalcik/Shutterstock.com*

## B – Existing detection methods

- ▶ For partially known GMO
  - ▶ New sequencing techniques coupled with
    - Molecular methods [Fraiture *et al.*, 2017, 2018]
    - Bioinformatics/Statistics [Willems *et al.*, 2016]
  - ▶ DNA walking [Fraiture *et al.*, 2015, 2018]
- ▶ For unknown GMO
  - ▶ No method available so far



## C – State of the art

### ► Current detection limits

	Detection of known and partially known GMO		Detection of unknown GMO	
	Prokaryotes	Eukaryotes	Prokaryotes	Eukaryotes
Intergenic sequences	✓	✓	✗	✗
Truncated gene	✓	✓	~	~
Fused gene	✓	✓	~	~
Insertion/deletion in a gene	✓	✓	✗	✗
% of point mutations $\geq 9\%$	✓	✓	✗	✗
% of point mutations $< 9\%$	✓	✓	✗	✗

1. Introduction
- 2. Objectives**
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## A – General information

- ▶ No method available for the detection of unknown GMO
- ▶ Creation of a method to address this issue
  - ▶ **DUGMO** : Detection of Unknown Genetically Modified Organism [Hurel *et al.*, BMC Bioinformatics 2020]  
<https://github.com/ANSES-Ploufragan/DUGMO>
- ▶ Basic idea of the method
  - ▶ Identify the vocabulary differences between the host genome and the insert

## B – General principle of DUGMO

- ▶ **Particularity** : use the CDS of the host genome
- ▶ Specific genomic vocabulary
  - ▶ Species-specific
  - ▶ Composed of words

### Nucleotidic sequence

**Species 1**

GTCGG**GTCG**ACGTCGGTCGTGTCGAGTCGG



4-letter words

## B – General principle of DUGMO

- ▶ **Particularity** : use the CDS of the host genome
- ▶ Specific genomic vocabulary
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  - ▶ Composed of words

### Nucleotidic sequence

**Species 1**      GTCGGGTCGACGTCGGTCGTGTCGAGTCGG

Over-represented 4 letter-words

**Species 2**      AGGCTCAGCTGAGCTTCA**GTCGG**TGTACTAGC

## C – Steps in DUGMO

### 1. Data preparation

- Cleaning, assembly, annotation pipeline
- Sorting of the CDSs in the sample
- Filtering of the databank of known GMOs

2. Characterisation of the « host » genome CDSs based on their over-represented words

3. Calculation of the distances to support the comparison to the host genome CDSs

4. Design of a prediction model

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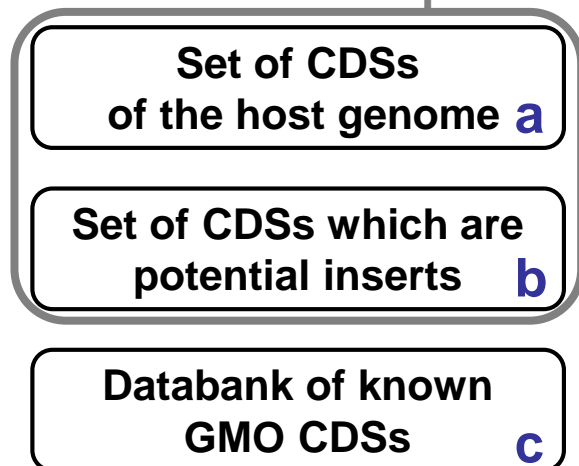
### 3. Calculation of the distances to support the comparison to the host genome CDSs

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## C – Steps in DUGMO

- ▶ Data preparation : cleaning/assembly/annotation, sorting and filtering

### Sample

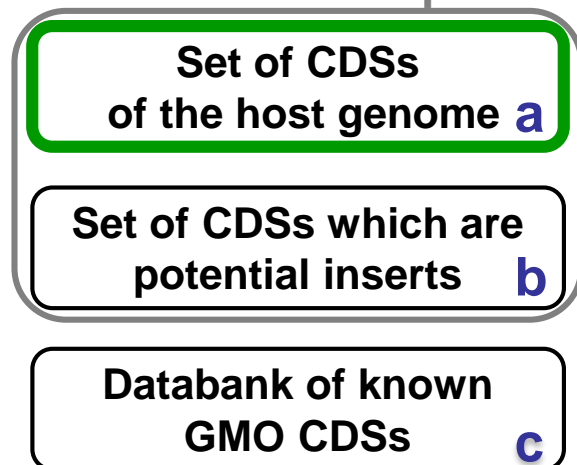


- ▶ Characterizes what we are not looking for
- ▶ Other CDSs, absent from the pangenome
- ▶ Non-species sequence variability
- ▶ Computation of distances to characterize :
  - ▶ the vocabulary of **a**
  - ▶ the vocabulary of **c**
  - ▶ the vocabulary of each sequence in the set **b**
- ▶ Design of a prediction model

## C – Steps in DUGMO

- ▶ Data preparation : cleaning/assembly/annotation, sorting and filtering

### Sample

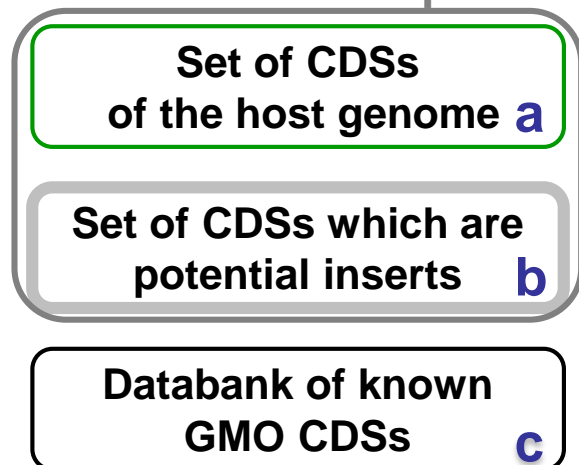


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- ▶ Data preparation : cleaning/assembly/annotation, sorting and filtering

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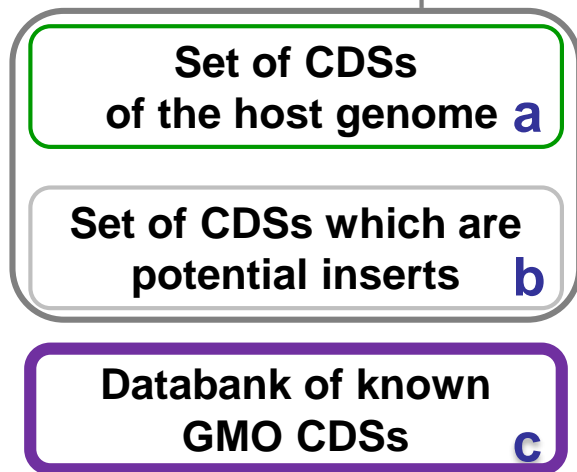


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- ▶ Data preparation : cleaning/assembly/annotation, sorting and filtering

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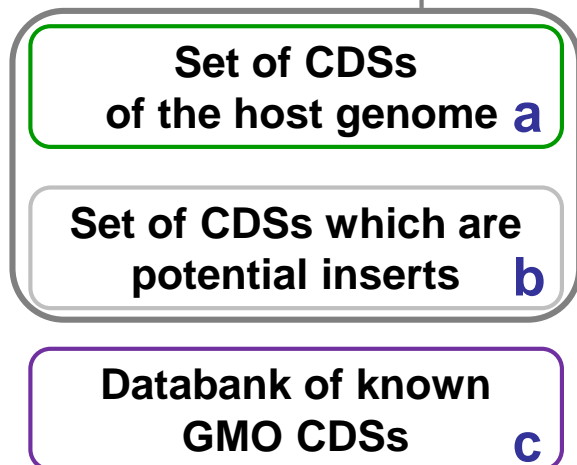


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## C – Steps in DUGMO

- ▶ Data preparation : cleaning/assembly/annotation, sorting and filtering

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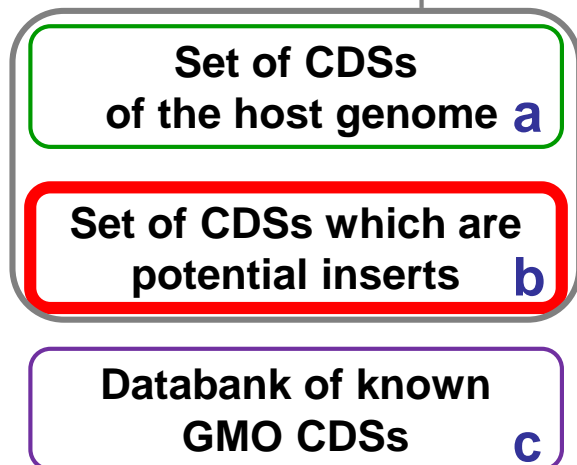


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## C – Steps in DUGMO

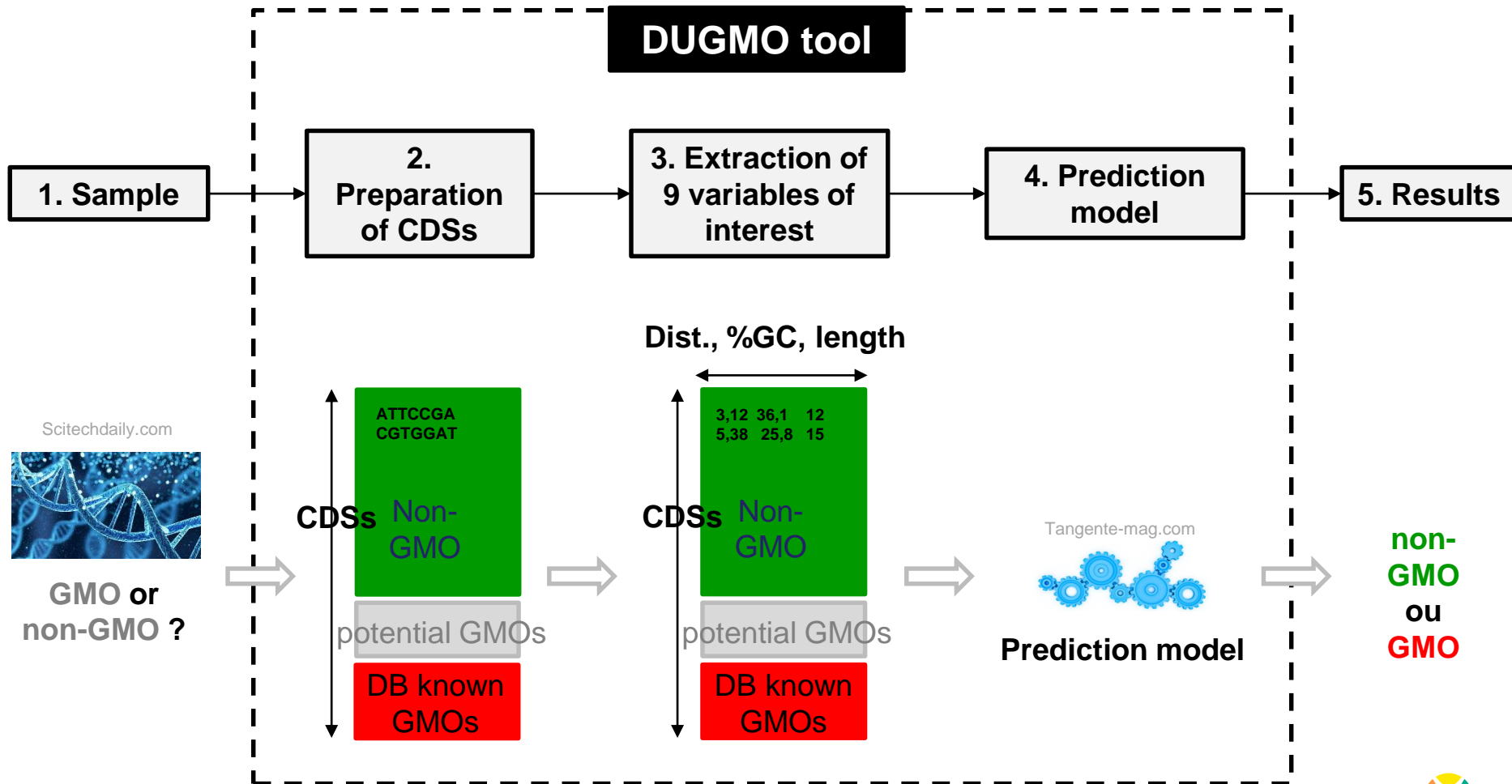
- ▶ Data preparation : cleaning/assembly/annotation, sorting and filtering

Sample



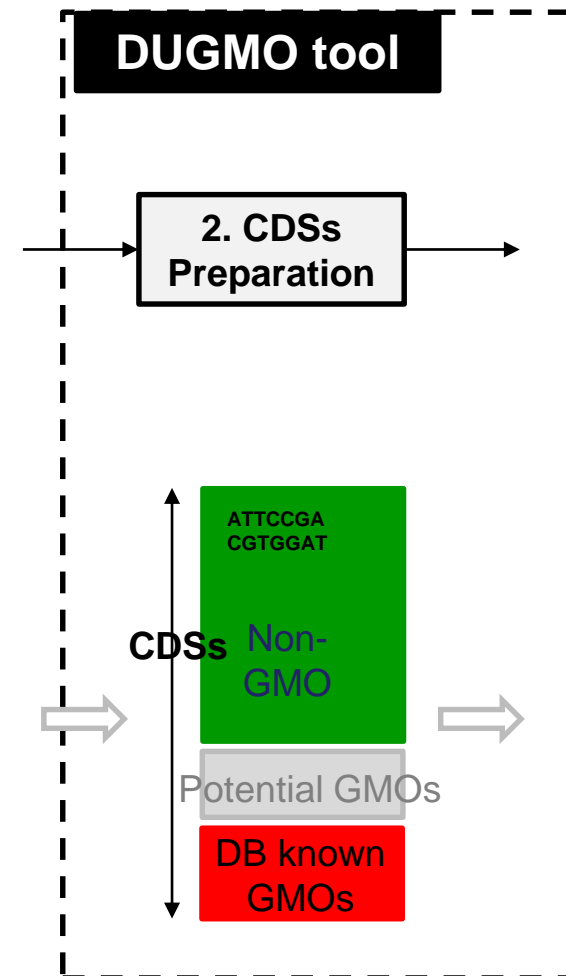
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- ▶ Computation of distances to characterize :
  - ▶ the vocabulary of **a**
  - ▶ the vocabulary of **c**
  - ▶ the vocabulary of each sequence in the set **b**
- ▶ Design of a prediction model => **GMO ?**

## C – Steps in DUGMO



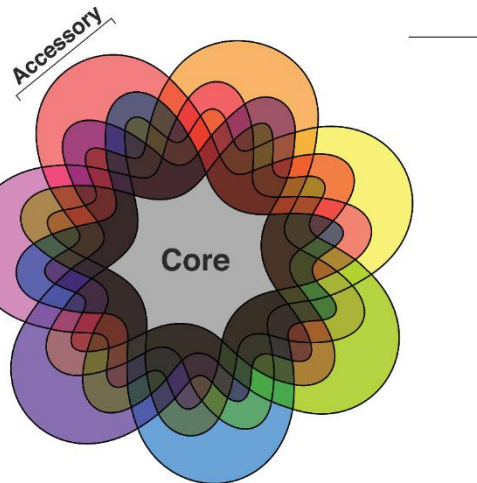
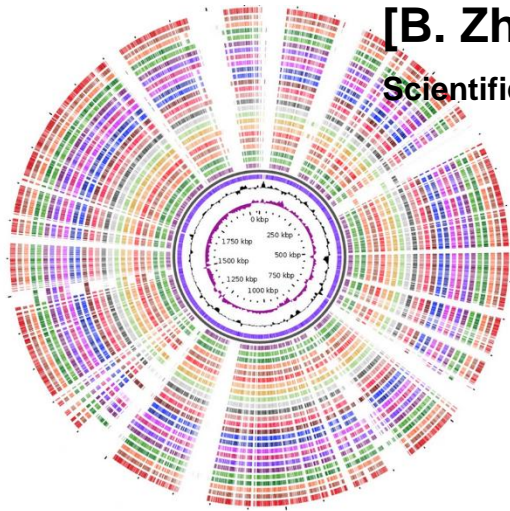


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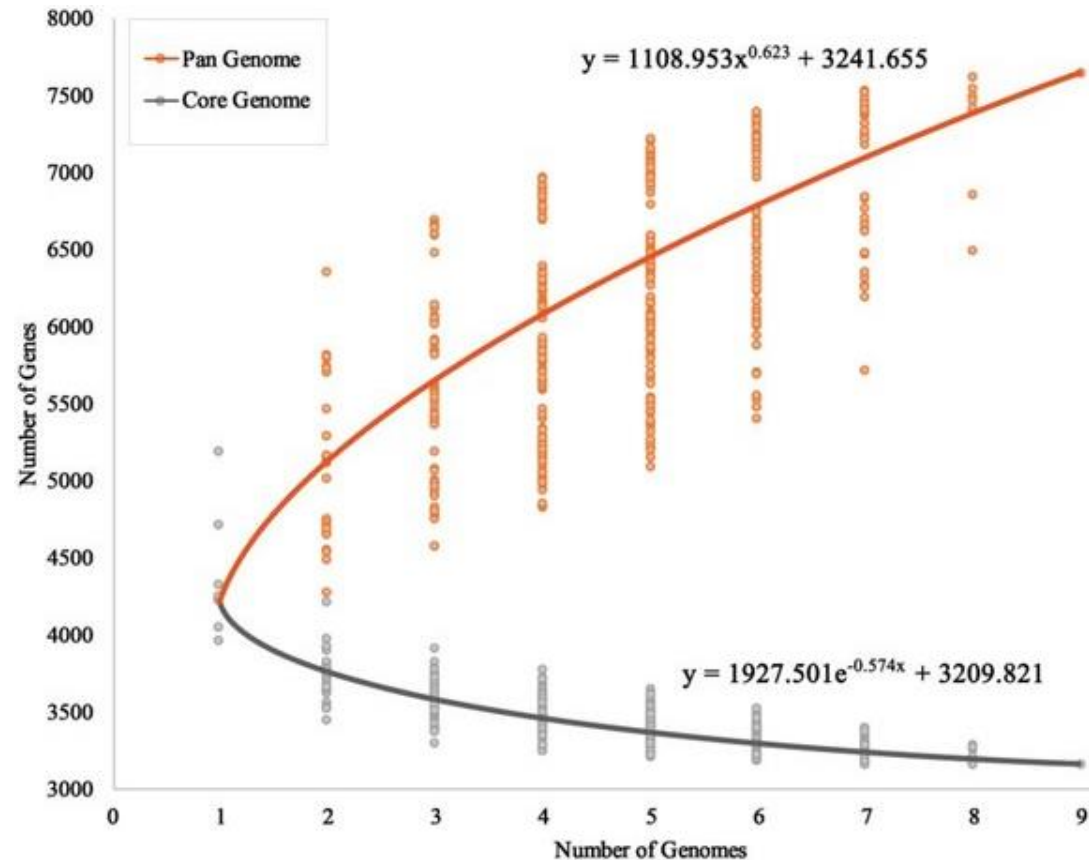


## A – Pangenome: wild-type and representative of species

[B. Zheng et al.  
ScientificReports 2017]



[CGP. McCarthy et al.  
Microbial genomics 2019]

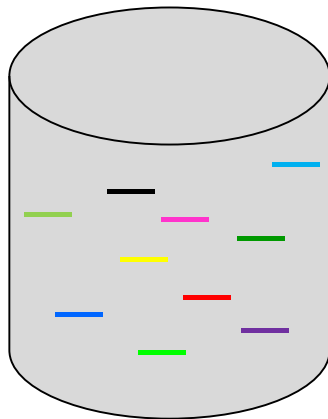


[D. Sinha et al.  
BMC genomics 2021]

## A – Data required as input to the software



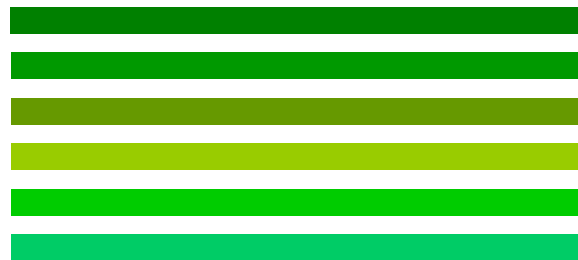
1. High throughput sequencing data



2. Reference genome (wild-type)



3. Pangenome (wild-type) and its associated CDSs



Species of the potentially GM bacterium

4. Databank of known GMO inserts

## B – Objectives

- ▶ Data cleaning, assembly, annotation of sequencing data
- ▶ Sorting of the sample CDSs
- ▶ Filtering of the database of known GMO inserts



High throughput  
sequencing data

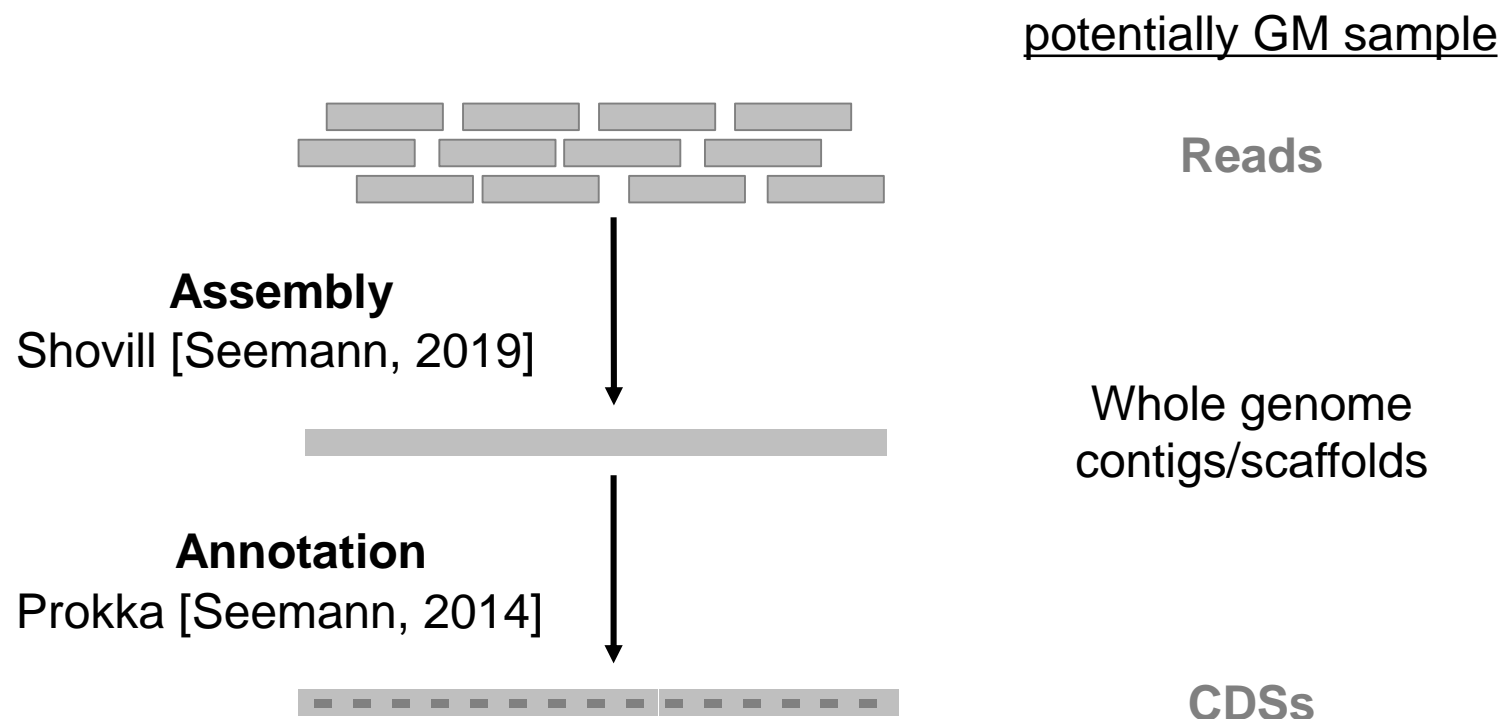
Cleaning, assembly,  
annotation then sorting

**CDSs related to the  
host genome**

CDSs which are  
potential GMO inserts

## B – Cleaning pipeline

- ▶ Assembly and annotation of high throughput sequencing data



## C – Sorting of the sample CDSs

- ▶ Step 1 : Comparison of the potential GMO CDSs with pangenome's CDSs

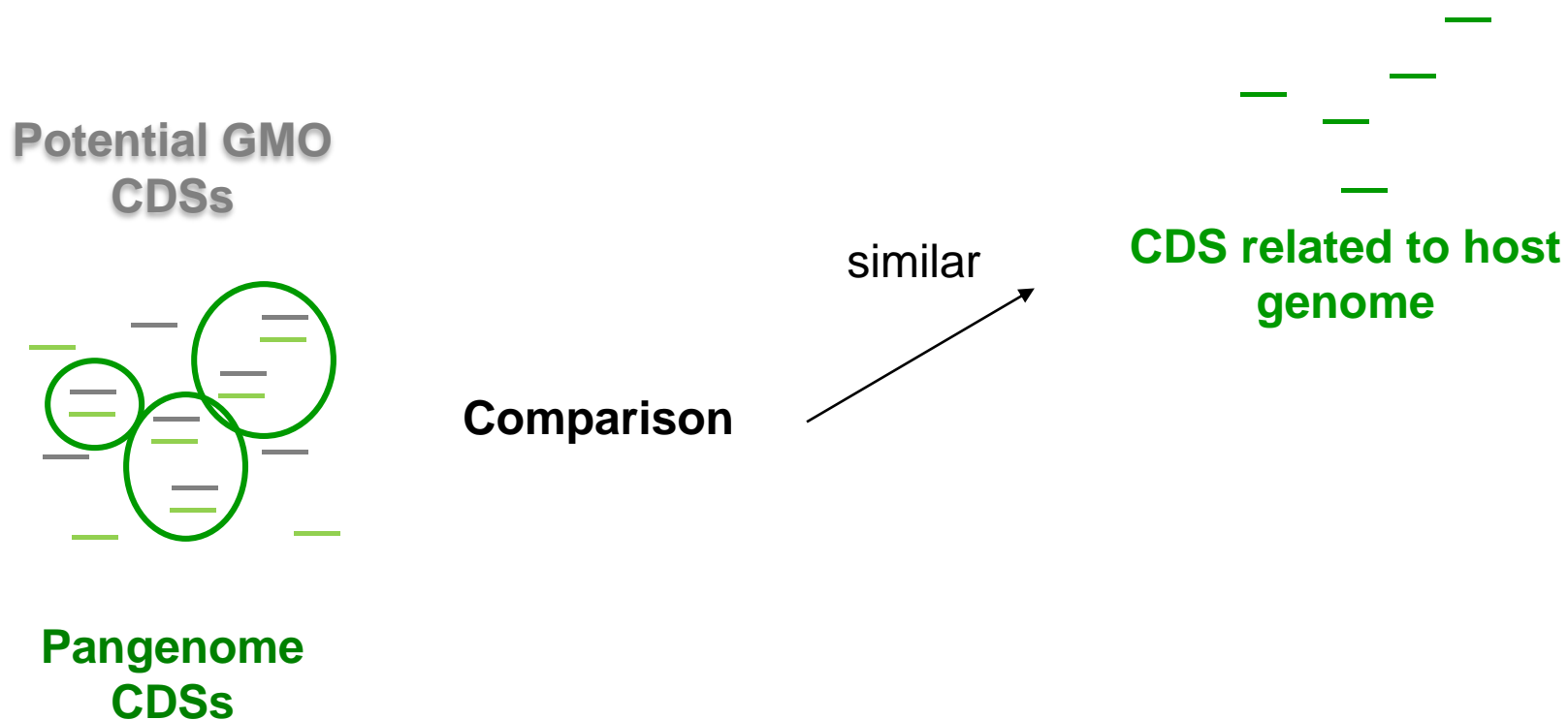
Potential GMO  
CDSs



Pangenome  
CDSs

## C – Sorting of the sample CDSs

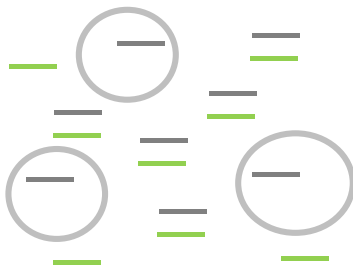
- ▶ Step 1 : Comparison of the potential GMO CDSs with pangenome's CDSs



## C – Sorting of the sample CDSs

- ▶ Step 1 : Comparison of the potential GMO CDSs with pangenome's CDSs

Potential GMO  
CDSs



Comparison

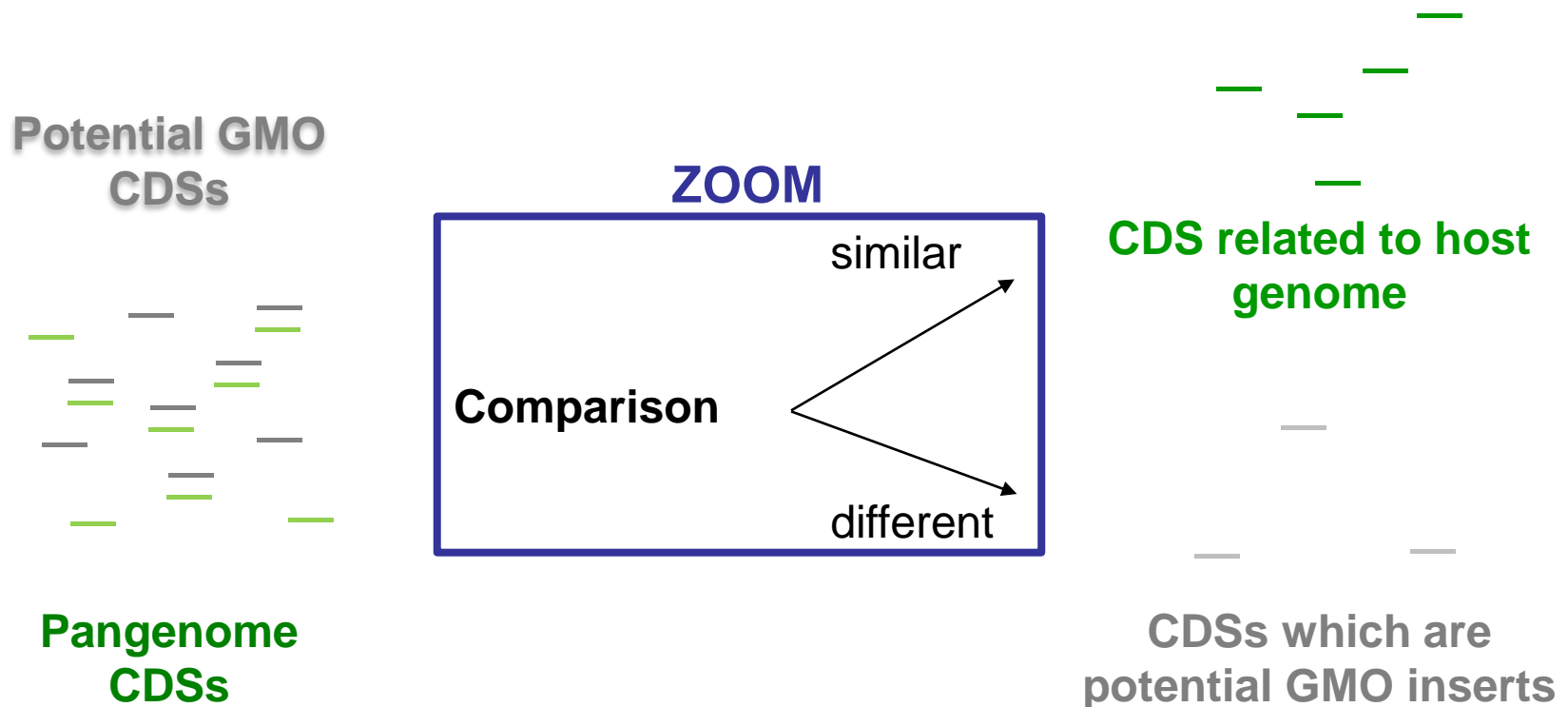
different

CDSs which are  
potential GMO inserts



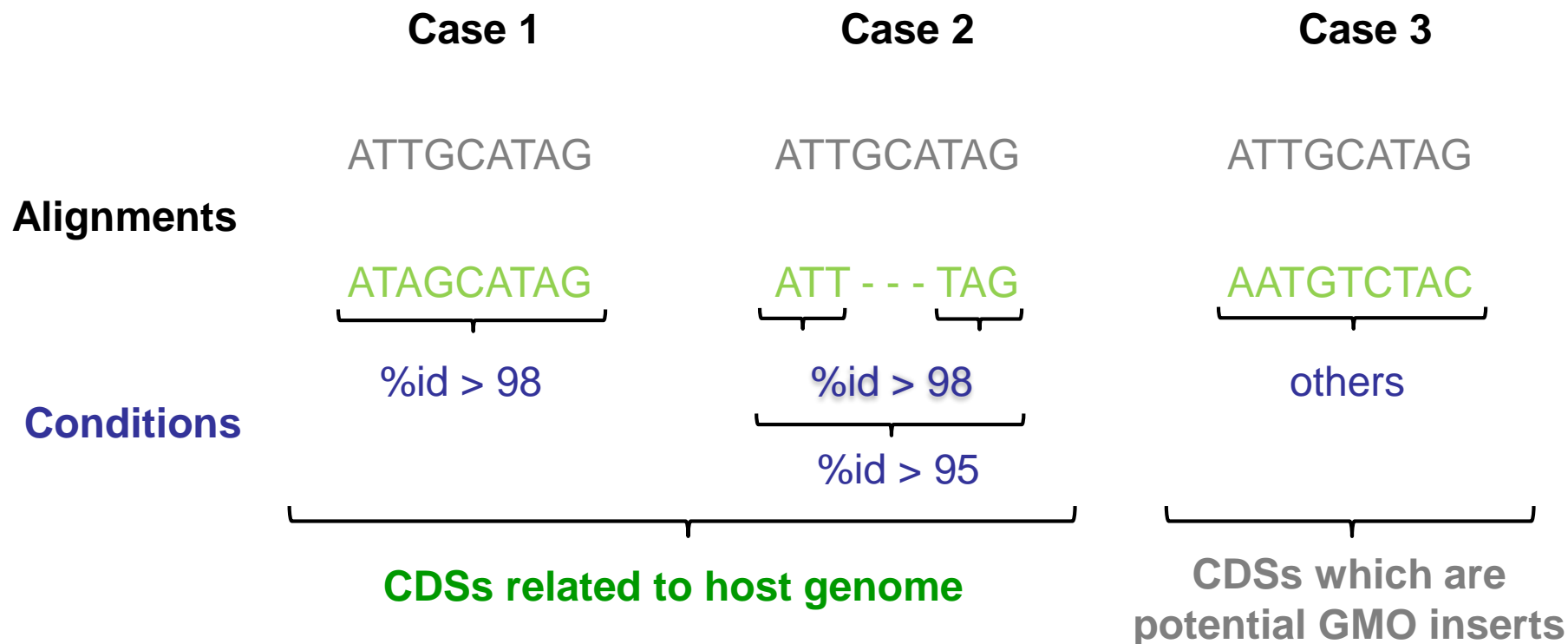
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## C – Sorting of the sample CDSs

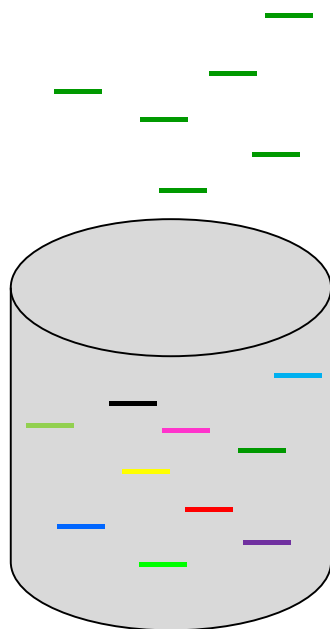
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## D – Filtering the GMO insert databank

- ▶ Using the host genome CDSs

**CDS related to the  
host genome**



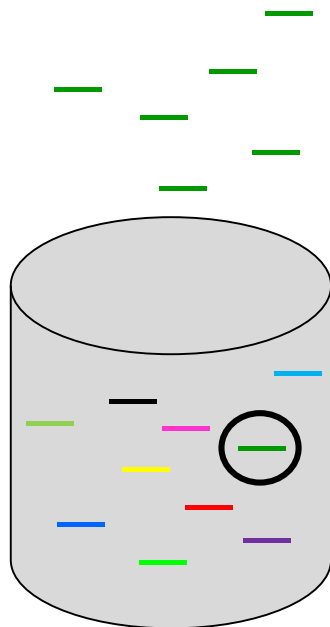
**Databank of known  
GMO inserts**

**Comparison criteria identical  
to step 1**

## D – Filtering the GMO insert databank

- ▶ Using the host genome CDSs

CDS related to the  
host genome



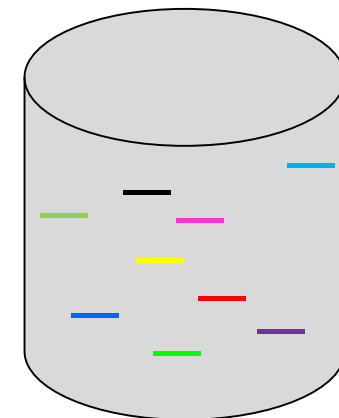
Databank of known  
GMO inserts

Comparison

similar

different

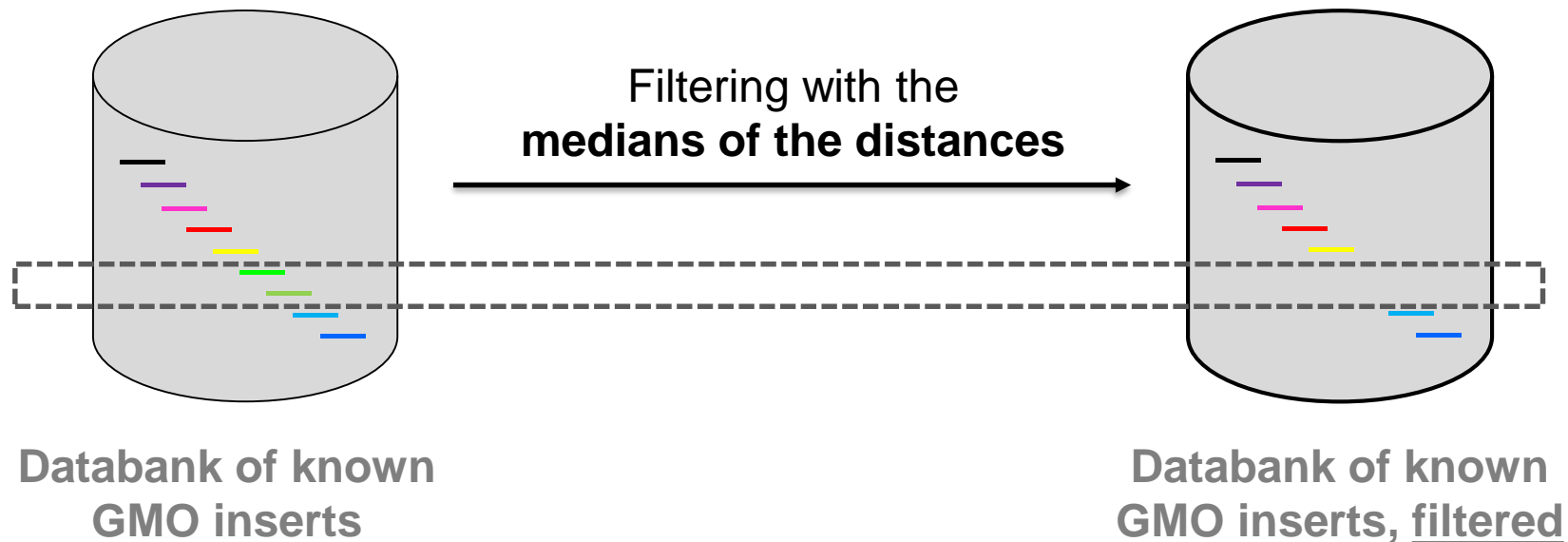
Discarded CDS



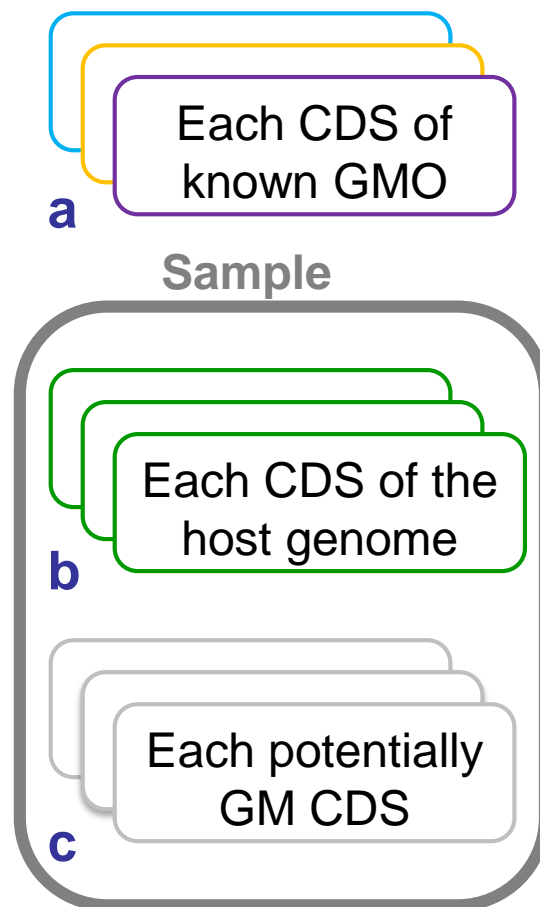
Databank of known  
GMO inserts, filtered



## D – Filtering the GMO insert databank



## C – Get the three needed distinct datasets

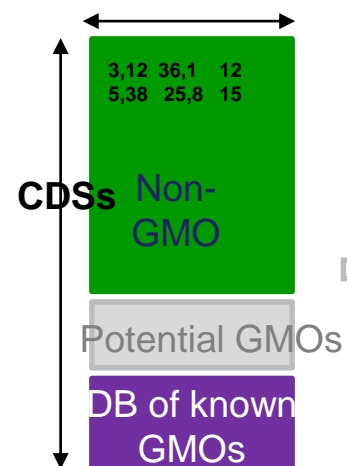


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## DUGMO tool

3. Extraction of  
the 9 variables  
of interest

Dist., %GC, length



## A – Objectives

- ▶ **Objective** : characterization of the host genome CDSs and of the two other CDS sets
- ▶ **Approach** : determine the best combinations of parameters
- ▶ **Purpose** : establish explanatory variables for the development of the prediction model



## B – Characterization of the genomic vocabulary

- ▶ **R'MES** [Schbath, S. and Hoebeke, M. 2011]: Searches for the exceptional words in a sequence
    - ▶ Determine the number of occurrences of each word and its exceptional character
    - ▶ It defines a set of words that are over-represented in the host genome
- R'MES**
- ▶ Three distance formula and two types of calculations
    - ▶ Euclidean distance
    - ▶ Kullback-Leibler distance [Trifonov et Rabadan, 2010]
    - ▶ Bray-Curtis distance

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**R'MES**

- ▶ Three distance formula and two types of calculations
  - ▶ Euclidean distance
  - ▶ Kullback-Leibler distance [Trifonov et Rabadan, 2010]
  - ▶ Bray-Curtis ~distance (dissimilarity measurement)

## B – Characterization of the genomic vocabulary

- ▶ Type of distance calculation named « in Frequencies »
  - ▶ Concatenation of the third codon positions into a new sequence

	<u>Word size</u>	<u>Running R'mes</u>
AGTACGTCAGGTAGTATCCAGCTAATG	27	impossible
TGATTCGAG	9	fast

- ▶ 10% of over-represented words
- ▶ Type of distance calculation named « in Proportions »
  - ▶ Whole CDS
  - ▶ All words

## B – Characterization of the genomic vocabulary

- ▶ Type of distance calculation named « in Frequencies »
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AGTACGTCAGGTAGTATCCAGCTAATG

TGATTGAG

Arginine

CGU

CGC

CGA

CGG

- ▶ 10% of over-represented words
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- ▶ 10% of over-represented words
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  - ▶ Whole CDS
  - ▶ All words

## C – Combinations of parameters

- ▶ List of tested parameters :
  - ▶ Word sizes
  - ▶ Order of the Markov model
  - ▶ Percentage of over-/under-represented words
  - ▶ Concatenation of the third codon positions

## D – Explanatory variables retained in DUGMO for one CDS

- ▶ Bray-Curtis distances
  - ▶ « In frequencies »
    - **F L9M7** : Word size 9 and Markov model order 7
  - ▶ « In proportions »
    - **P L3M1** : Word size 3 and Markov model order 1
    - **P L4M2** : Word size 4 and Markov model order 2
  
- ▶ Average exceptionality scores in the host genome for L4M2 et L9M7
  
- ▶ Count density per nucleotide for 4-letter and 9-letter words
$$\frac{\text{Sum of all word counts}}{\text{CDS length}}$$
  
- ▶ Percentage of GC
  
- ▶ Length

## D – Explanatory variables retained in DUGMO for one CDS

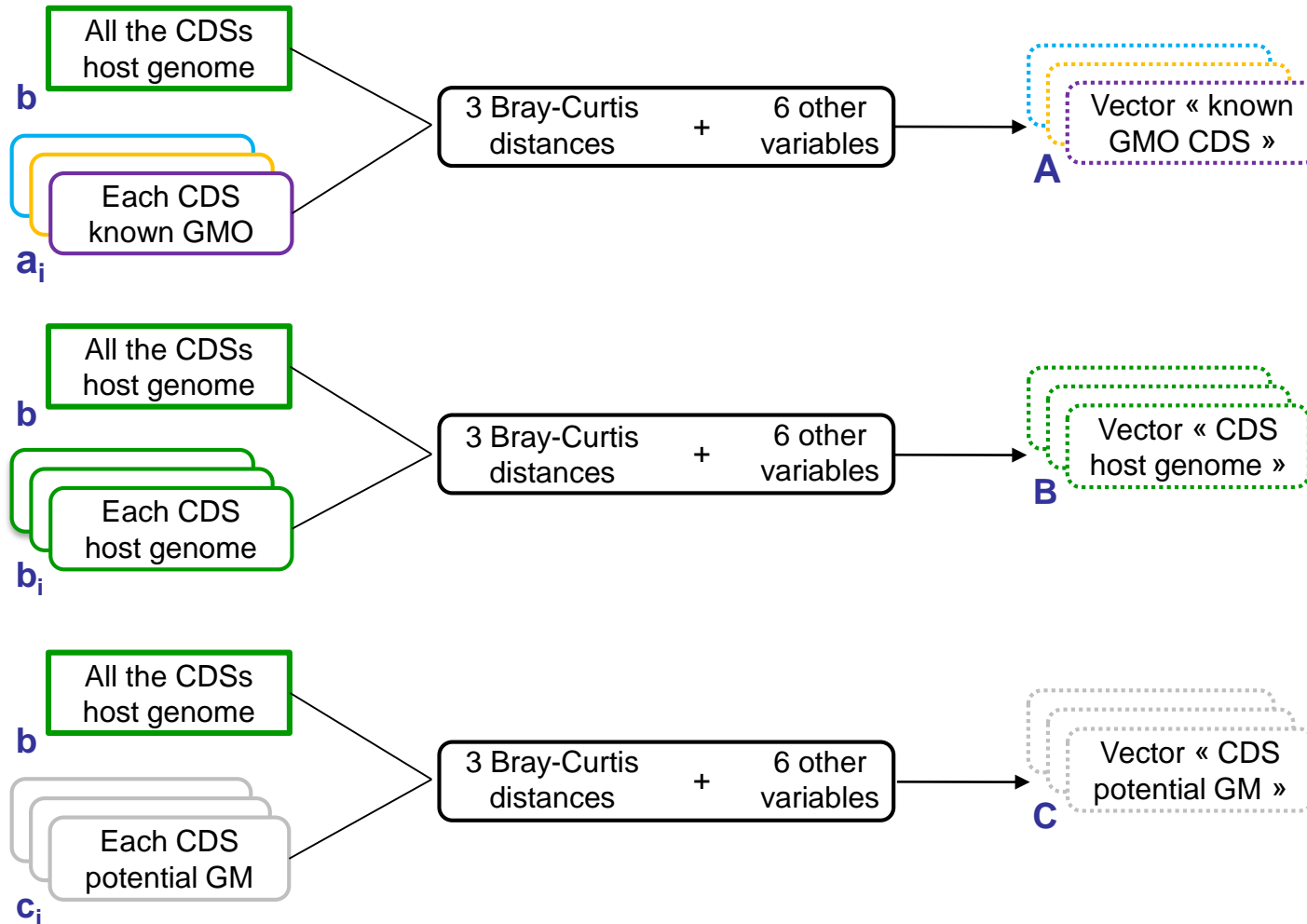
- ▶ Bray-Curtis distances
  - ▶ « In frequencies »
    - **F L9M7** : Word size 9 and Markov model order 7
  - ▶ « In proportions »
    - **P L3M1** : Word size 3 and Markov model order 1
    - **P L4M2** : Word size 4 and Markov model order 2
- ▶ Average exceptionality scores provided by R'MES in the host genome for L4M2 and L9M7
- ▶ Count density per nucleotide for 4-letter and 9-letter words

$$\frac{\text{Sum of all word counts}}{\text{CDS length}}$$

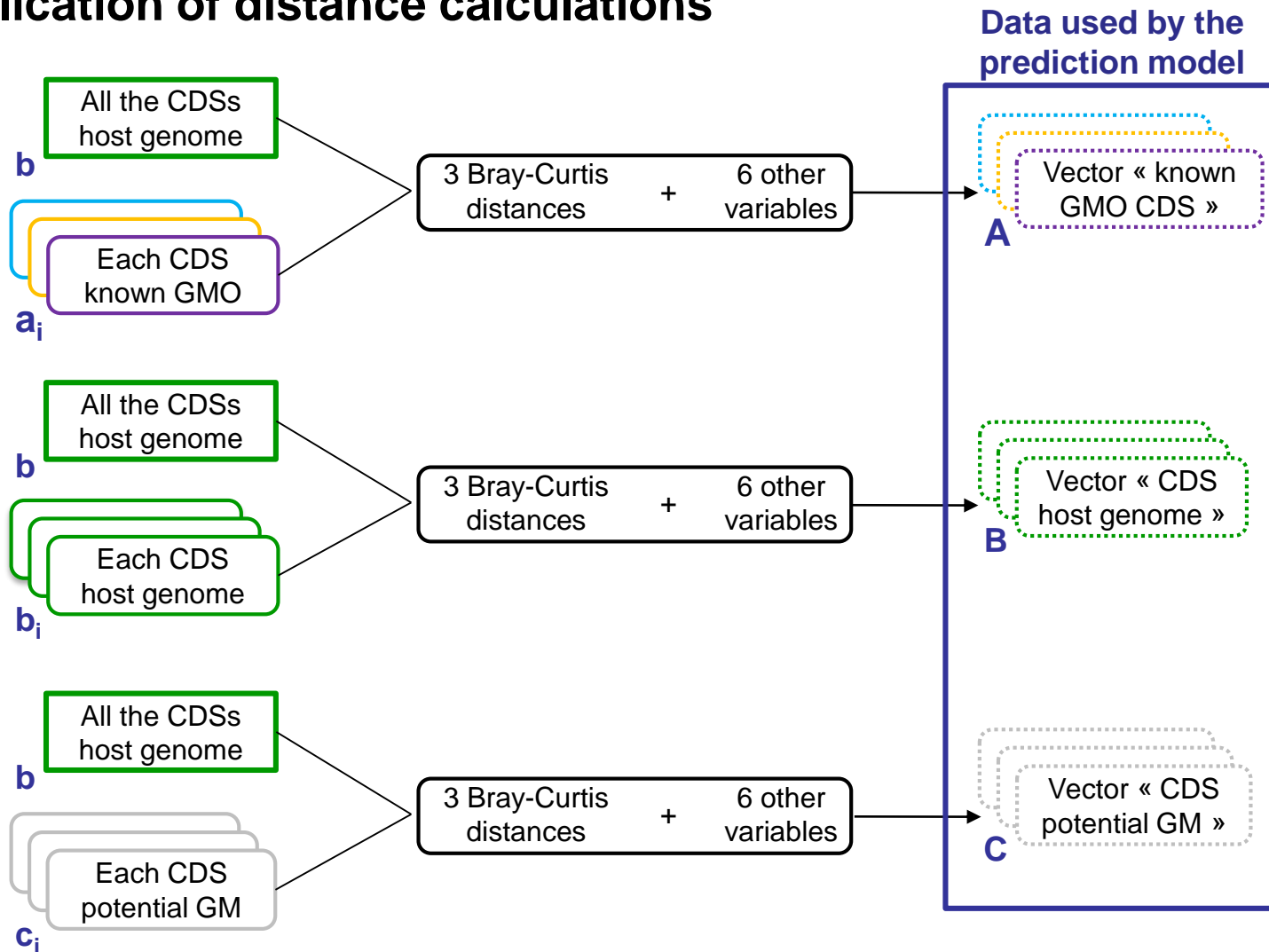
- ▶ Percentage of GC
- ▶ Length



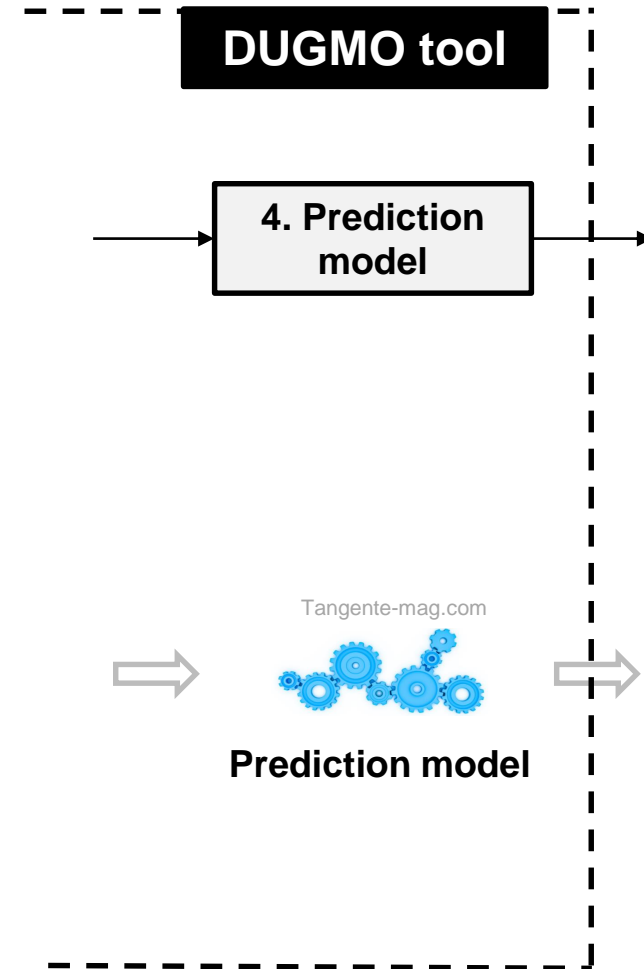
## E – Application of distance calculations



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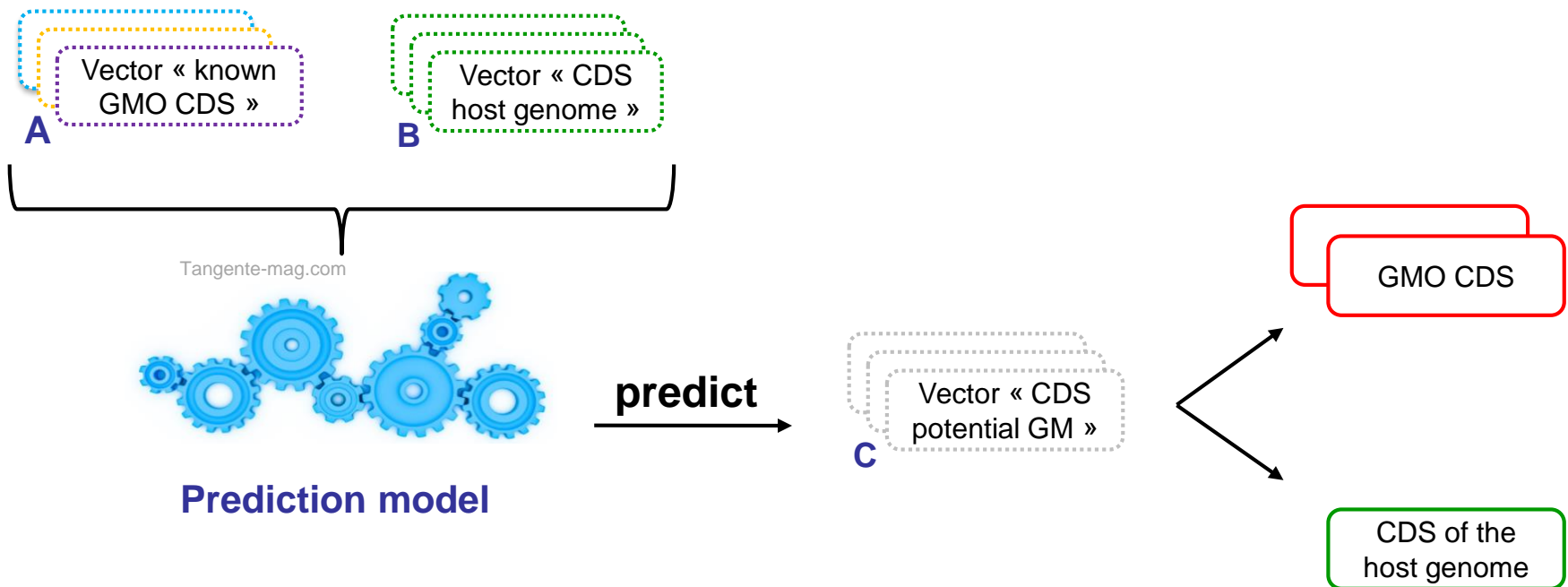


1. Introduction
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3. Préparation des données
4. Calcul de distances
- 5. Design of the prediction model**
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## A – Objectives

- ▶ **Approach** : use of Machine Learning methods
- ▶ **Purpose** : predict proven CDSs of GMO inserts



## B – Machine Learning

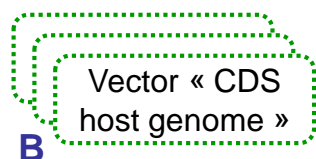
### ▶ 12 tested methods

- |  |                    |
|--|--------------------|
| ▶ Logit : generalized linear model                   | parametric         |
| ▶ StepLDA : linear discriminant analysis             |                    |
| ▶ StepQDA : quadratic discriminante analysis         |                    |
| ▶ Plsda : partial least squares regression           |                    |
| ▶ NN : neural networks                               | non parametric     |
| ▶ SvmRadial : support vector machines                | including decision |
| ▶ KNN : K nearest neighbors                          | trees              |
| ▶ <u>C5.0 : classification algorithm</u>             |                    |
| ▶ <u>Rpart : recursive partitionig trees</u>         |                    |
| ▶ <u>RF : random forests</u>                         |                    |
| ▶ <u>Treebag : classification trees with bagging</u> |                    |
| ▶ <u>Xgboost : extreme gradient boosting</u>         |                    |

## C – Used data



▶ CDSs of the GMO insert databank after filtering



▶ CDSs related to the host genome

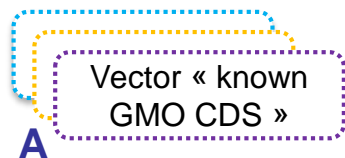


▶ potential GMO CDSs

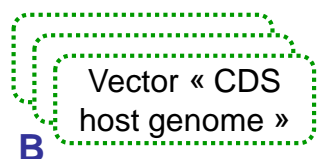
**Training data**

**Predictive data**

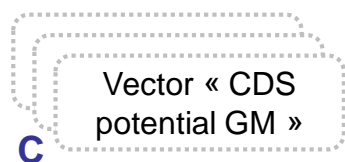
## C – Used data



▶ CDSs of the GMO insert databank after filtering



▶ CDSs related to the host genome



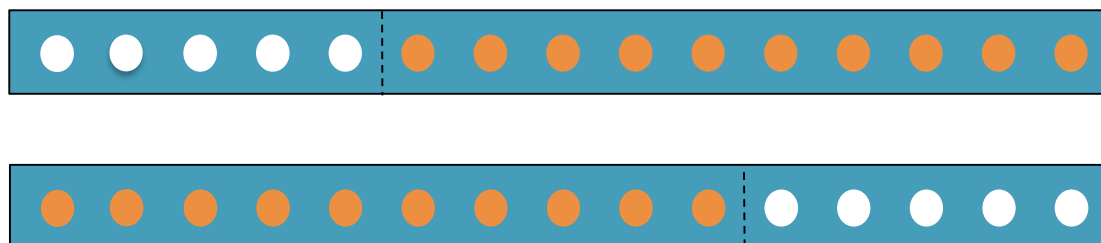
▶ Potential GMO CDSs

Training data

Predictive data

## D – Comparison of the tested methods

- ▶ Centered, reduced, and stratified data
- ▶ Optimisation of the parameters of each method in 10-fold cross-validation
- ▶ 2-fold cross-validation comparison of methods



2-fold cross-validation



## E – Selection criteria

### ► Confusion matrix

		Predictions of a model	
		GMO	Non-GMO
Real data	GMO	True positives	False negatives
	Non-GMO	False positives	True negatives

► **Specificity : ++**

► **False positive rate: --**

► **Sensitivity : ++**

► **False negative rate : +++**

## F – Final choice

- ▶ Union of the results of two methods
  - ▶ **RF** : Random Forests
  - ▶ **Logit** : Generalized linear model

Results for prediction data			
	Logit	RF	Union of RF and Logit
<b>False negative rate</b>	0.04	0.01	0.01
<b>Specificity</b>	0.94	0.98	0.99
<b>Sensitivity</b>	0.95	0.98	0.99
<b>False positive rate</b>	0.05	0.01	0.09

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**DUGMO tool**

**5. Results**

**GMO**  
or  
**non-  
GMO**

## A – Tested data

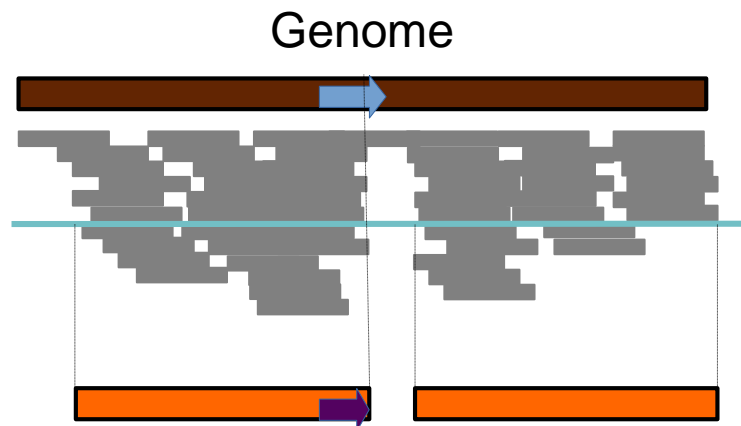
- ▶ **1 wild-type** genome and **1 GM** genome of *B. subtilis* bacterium
- ▶ **3 GM** genomes of *E. coli* bacterium
- ▶ **6 wild-type** genomes of bacteria
  - ▶ *Campylobacter jejuni*
  - ▶ *Lactococcus lactis*
  - ▶ *Listeria monocytogenes*
  - ▶ *Mycobacterium tuberculosis*
  - ▶ *Staphylococcus aureus*
  - ▶ *Salmonella Typhimurium*
- ▶ **42 synthetic GM** genomes
  - ▶ Combinations : 6 wild-type bacteria + 7 exogenous genes

## B – Global results

- ▶ With the 2 *B. subtilis* genomes
  - ▶ WT: No insert ✓
  - ▶ GM: 25 detected inserts +  
12 false negatives (maximum) ✓
  
- ▶ With the 3 GM genomes of *E. coli*
  - ▶ 3 known inserts are detected ✓
  - ▶ 1 false positive ✗ → ✓
  
- ▶ With the 48 synthetic WT genomes
  - ▶ 47: No insert ✓
  - ▶ 1 false positive (*M. tuberculosis*) ✗ → ✓
  
- ▶ With the 48 synthetic GM genomes
  - ▶ 47: insert found ✓
  - ▶ 1 false negative (*S. aureus* including a *C. jejuni* gene) ✗

## C – GM *Escherichia coli*

- ▶ In the genome modified with a gene from *S. pyogenes* : 1 false positive
  - ▶ *arlS* gene
  - ▶ **Truncated gene because of the assembly**



### Legend

Original genome, gene

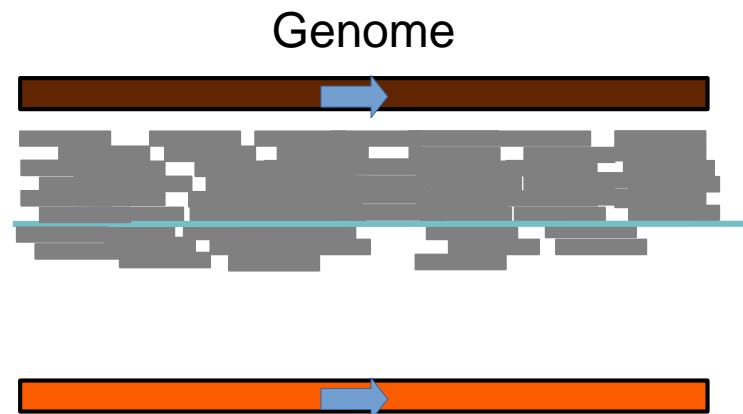
Reads

Minimal coverage depth (60)

assembled genome, truncated  
gene

## C – GM *Escherichia coli*

- ▶ In the genome modified with a gene from *S. pyogenes* : ~~1 false positive~~
- ▶ Solution : **minimal coverage depth of 60** (bacteria)



### Legend

original genome, gene

Reads

Minimal coverage depth (60)

assembled genome

## D – Wild-type *Mycobacterium tuberculosis*

- ▶ One false positive detected by DUGMO: **labelled GM while it is not**

CDS in the potential  
GM genome

—  
—

Comparison  
%id < 95%

different

False positive

CDS in the  
pangénome

MEGABLAST

NCBI  
WGS

No other similar gene



Horizontal gene transfert ?

Phylum

Actinomycetota

Order

Actinomycetes

Sub-Order

Corynebacteriales

Family

Mycobacteriaceae

Genus

Mycobacterium

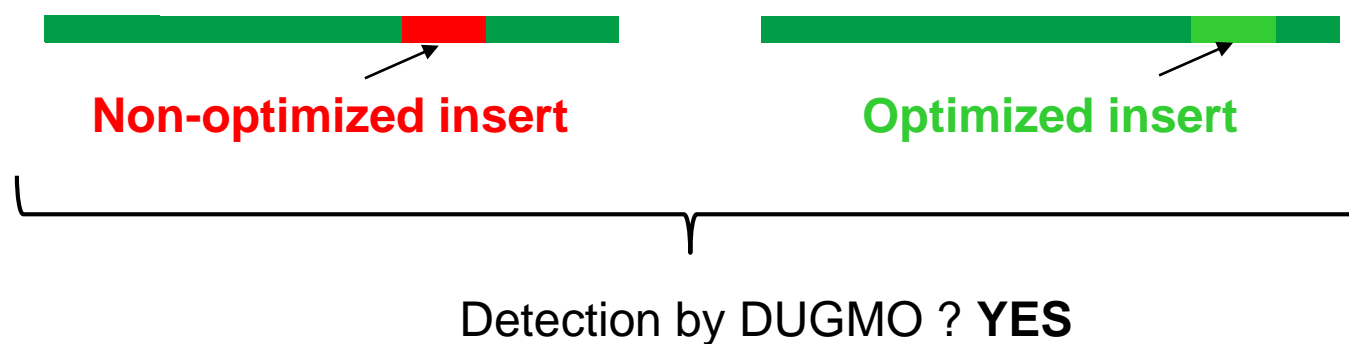


## E – Synthetic data

- ▶ Objectives

- ▶ Test the sensitivity of the method to **dicodon optimization**

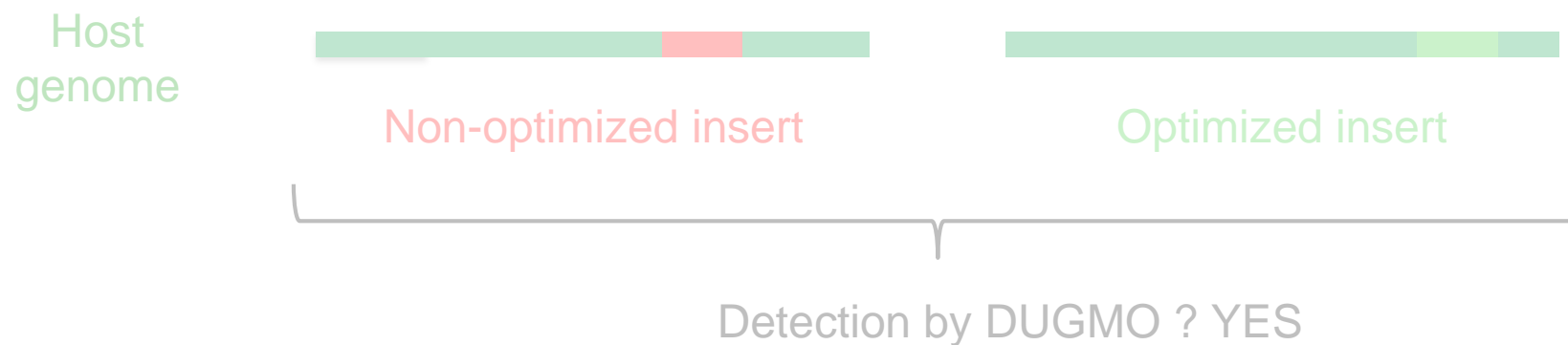
Host  
genome



## E – Synthetic data

### ▶ Objectives

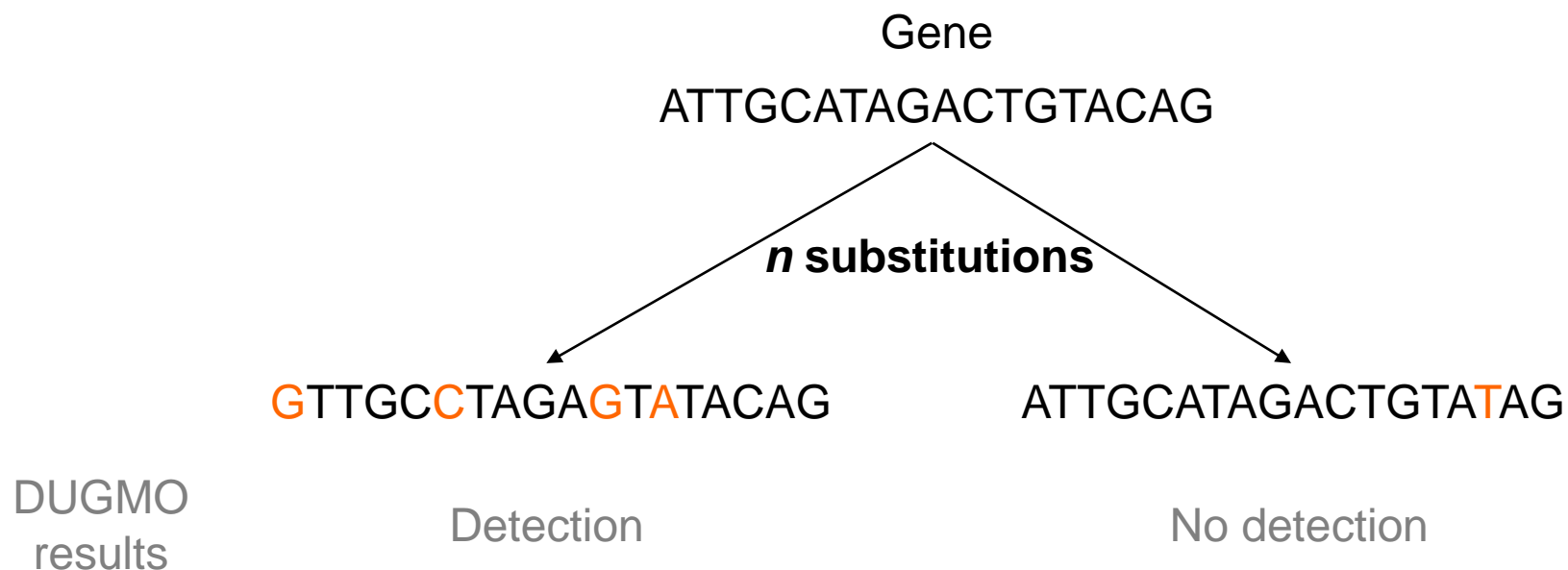
- ▶ Test the sensitivity of the method to dicodon optimization



- ▶ Test the **detection threshold** of the method

## E – Synthetic data

- ▶ Generation of **mutations in a wild-type gene of *B. subtilis***



- ▶ Results : **substitution rate  $\geq 9\%$**  → **detection of the GM gene**

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## C – Summary: before

	Detection known or partially known GMO		Detection unknown GMO	
	Prokaryotes	Eukaryotes	Prokaryotes	Eukaryotes
Intergenic sequence	✓	✓	✗	✗
Truncated gene	✓	✓	~	~
Fused gene	✓	✓	~	~
Insertion/deletion in a gene	✓	✓	✗	✗
% of point mutations $\geq 9\%$	✓	✓	✗	✗
% of point mutations $< 9\%$	✓	✓	✗	✗

## C – Summary: after

	Detection known or partially known GMO		Detection unknown GMO	
	Prokaryotes	Eukaryotes	Prokaryotes	Eukaryotes
Intergenic sequences	✓	✓	✗	✗
Truncated gene	✓	✓	✓	⌚
Fused gene	✓	✓	✓	⌚
Insertion/deletion in a gene	✓	✓	✓	⌚
% of point mutations $\geq 9\%$	✓	✓	✓	⌚
% of point mutations $< 9\%$	✓	✓	✗	✗

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## A – General

- ▶ Adapt the method for application to other organisms



tokopedia.com Tetra Import Glowfish

- ▶ Possibility to provide assembly data as input (Illumina, Pacbio)

GloFish®



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Federal Office of  
Consumer Protection  
and Food Safety



## ► Funders

Anses



Région Bretagne

# Thank you for your attention

<https://github.com/ANSES-Ploufragan/DUGMO>

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Answers reflect my personal opinion, they are not the expression of the ANSES opinion

# 18 months post-doc position in bioinformatics

## DUGMO for Eukaryotic genomes

Conditions:

- out of France for 18 months from May 2020
- work in a P2+ lab

Contact: [fabrice.touzain@anses.fr](mailto:fabrice.touzain@anses.fr)

<https://github.com/ANSES-Ploufragan/DUGMO>